

Use of Protein Cross-Linking and Radiolytic Labeling To Elucidate the Structure of PsbO within Higher-Plant Photosystem II

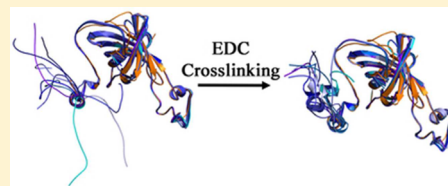
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ABSTRACT: We have used protein cross-linking with the zero-length cross-linker 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide, and radiolytic footprinting coupled with high-resolution tandem mass spectrometry, to examine the structure of higher-plant PsbO when it is bound to Photosystem II. Twenty intramolecular cross-linked residue pairs were identified. On the basis of this cross-linking data, spinach PsbO was modeled using the *Thermosynechococcus vulcanus* PsbO structure as a template, with the cross-linking distance constraints incorporated using the MODELLER program. Our model of higher-plant PsbO identifies several differences between the spinach and cyanobacterial proteins. The N-terminal region is particularly interesting, as this region has been suggested to be important for oxygen evolution and for the specific binding of PsbO to Photosystem II. Additionally, using radiolytic mapping, we have identified regions on spinach PsbO that are shielded from the bulk solvent. These domains may represent regions on PsbO that interact with other components, as yet unidentified, of the photosystem.



Photosystem II (PS II) is a light energy-driven water-plastoquinone oxidoreductase. Excitation energy from the light-harvesting apparatus (phycobilisomes in cyanobacteria and light-harvesting chlorophyll proteins in green plants and algae) is transferred to the reaction center of the photosystem. Initial charge separation occurs between Chl_{D1} and Pheo_{D1} , which yields $\text{Chl}_{\text{D1}}^+\text{Pheo}_{\text{D1}}^-$.¹ This charge-separated state is stabilized by rapid electron transfer from $\text{Pheo}_{\text{D1}}^-$, first to Q_A and then to Q_B . The accumulation of two electrons on Q_B leads to protonation and its subsequent release as plastoquinol. Chl_{D1}^+ is reduced by P_{D1} , and P_{D1}^+ is reduced by Y_z , the residue D1:¹⁶¹Y, yielding $\text{Y}_\text{z}^\bullet$ with the release of a proton. The subsequent reduction of $\text{Y}_\text{z}^\bullet$ by the Mn_4CaO_5 cluster occurs via proton-coupled electron transport and leads to the accumulation of an oxidizing equivalent at the oxygen-evolving site, the Mn_4CaO_5 metal center. The accumulation of four oxidizing equivalents ultimately leads to the oxidation of two water molecules and the release of dioxygen.^{2,3}

Cyanobacterial PS II contains at least 20 subunits, of which at least 17 are intrinsic membrane proteins.⁴ A subset of these components (D1 , D2 , CP43 , CP47 , the α and β subunits of cytochrome b_{559} , and the PSBL protein) is required for oxygen evolution and accumulation of the photosystem in thylakoid membranes. Genetic or biochemical removal of any of these protein subunits leads to loss of functional PS II. In addition, 10 low-molecular mass intrinsic proteins are present; however, their functions remain obscure. PS II complexes containing only these intrinsic proteins evolve oxygen at low rates and require very high

concentrations of calcium and chloride.^{5,6} At presumptive physiological concentrations of these cofactors, efficient oxygen evolution requires the extrinsic components of the photosystem. These include PsbO, which is uniformly present in all oxygenic organisms, and either PsbU, PsbV, CyanoQ and CyanoP (in the cyanobacteria), or PsbP, PsbQ, and PsbR (in green plants) (for reviews, see refs 7–12). PsbO plays an important role in the stabilization of the Mn_4CaO_5 cluster¹³ at low chloride concentrations and protects the metal cluster from exogenous reductants. In higher plants, it is required for PS II assembly^{7,14–17} and photosynthetic oxygen evolution. Interestingly, in cyanobacteria, this is not the case.⁵ In the absence of PsbO, *Synechocystis* can grow autotrophically and evolve oxygen, albeit at lower rates.¹⁸

Over the past 15 years, increasingly higher resolution crystal structures of thermophilic cyanobacterial PS II have become available.^{4,19–24} These studies have been critically important in improving our understanding of the molecular organization of the photosystem. Recently, a crystal structure of PS II from the red alga *Cyanidium caldarium*, at 2.76 Å resolution, has also become available.²⁵ This is, importantly, the first structure of PS II available from a eukaryote. It should be noted, however, that the red algae do not lie in the green plant lineage.

As no crystal structure for higher-plant PS II is available, the structure and interactions of the extrinsic proteins with the

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intrinsic components of the photosystem have been modeled by comparison to those of cyanobacterial PS II. It must be stressed, however, that the interactions of the PsbP and PsbQ proteins with PS II are not well understood. Additionally, strong biochemical evidence indicates the presence of a putative second copy of PsbO being associated with each PS II monomer in higher plants (for reviews, see refs 13 and 26). Protein cross-linking coupled with tandem mass spectrometry is a powerful technique for elucidating protein–protein interactions in multisubunit protein complexes.^{27,28} Recently, these methods have been used to investigate subunit–subunit interactions within PS II and PS I.^{29–32} Liu et al.³⁰ elucidated the structure of a PS II–phycobilisome–PS I megacomplex using dithiobis(succinimidylpropionate). Liu et al.,³¹ using the cross-linkers 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC, a zero-length cross-linker) and dithiobis(succinimidylpropionate), identified a binding domain for CyanoQ within *Synechocystis* PS II. Ido et al.³² have extended their earlier studies using the cross-linker EDC on higher-plant PS II. In this study, they identified interchain cross-linked products involving PsbP, PsbQ, and other PS II components (CP43, CP26, and PsbR). Ido et al.³² have positioned PsbQ on the periphery of the PS II complex in marked contrast to the position of CyanoQ proposed by Liu et al.³¹ Our laboratory has used the cross-linker bis-sulfosuccinimidyl suberate to examine the interaction of PsbP and PsbQ in higher-plant PS II.³³ Our results indicate that PsbP and PsbQ directly interact, with the location of PsbQ being consistent with the location hypothesized by either Liu et al.³¹ or Ido et al.³² It should be noted that the proposed location of PsbQ presented by Ido et al.³² and modified by Mummadisetti et al.³³ is similar to the location of PsbQ' in the *Cyanidium* PS II crystal structure.²⁵ Recently, models for the structural organization of the higher-plant extrinsic proteins in association with PS II have been presented.¹²

The radiolysis of water by synchrotron radiation produces •OH, which is capable of oxidatively modifying amino acid residues that are in contact with the bulk solvent. The modified residues can be identified using tandem mass spectrometry, and their location can be used to “footprint” protein–protein interactions within multisubunit protein complexes.^{34–36} This radiolytic mapping technique has been used to identify both surface and buried amino acid residues that are exposed to water.^{37–40} Recently, we used this technique to examine the interaction of PsbP and PsbQ in higher-plant PS II.³³ In this study, we identified domains on PsbP that appear to be shielded from the bulk solvent by intrinsic components of PS II and, possibly, by PsbR.

In this paper, we have used protein cross-linking coupled with tandem mass spectrometry to identify 20 intramolecular cross-linked products within the PsbO protein. These experiments allowed the modeling of the N-terminus of higher-plant PsbO, which contains a 10-amino acid extension when compared to cyanobacterial PsbO. This facilitated the localization of the two binding determinants for higher-plant PsbO that previously had been identified in this region.^{41,42} Radiolytic mapping was also used to oxidatively modify surface residues on PsbO that were in contact with water. Seventy-seven residues were identified. Interestingly, a large domain on the surface of PsbO was resistant to radiolytic modification, indicating that it was shielded from the bulk solvent. This domain appears to define buried regions of PsbO that are in contact with other, as yet unidentified, components of PS II.

MATERIALS AND METHODS

PS II Membrane Isolation and Protein Cross-Linking. PS II membranes were isolated by the method of Berthold et al.⁴³ from market spinach. Chl was determined by the method of Arnon.⁴⁴ After isolation, the PS II membranes were suspended at a concentration of 2 mg of chl/mL in 50 mM Mes-NaOH (pH 6.0), 300 mM sucrose, 15 mM NaCl (SMN) buffer and frozen at -80°C until they were used. Protein cross-linking was performed using the zero-length cross-linker EDC. In our experiments, PS II membranes were suspended at a chl concentration of 200 $\mu\text{g}/\text{mL}$ in 25% glycerol, 10 mM MgCl_2 , 5 mM CaCl_2 , and 50 mM MES-NaOH (pH 6.0) buffer. The membranes were then treated with 6.25 mM EDC and 5 mM *N*-hydroxysulfosuccinimide for 2 h at room temperature in the dark. The reaction was quenched by bringing the reaction mixture to 100 mM ammonium bicarbonate and incubating the mixture for 20 min at room temperature. The membranes were harvested by centrifugation for 25 min at 38000g, and the final pellet was resuspended in 1.0 M NaCl in SMN buffer for 1 h at 4°C to release the PsbP and PsbQ subunits. The PsbP- and PsbQ-depleted PS II membranes were washed with SMN and then treated with 1.0 M CaCl_2 in SMN buffer to release PsbO. After centrifugation to remove the PsbO-depleted membranes, the supernatant from the CaCl_2 wash (which contains PsbO) was dialyzed overnight against 10 mM Mes-NaOH (pH 6.0) using a 6–8 kDa cutoff membrane (Spectrum Laboratories, Inc.) and was concentrated by ultrafiltration using a 10 kDa cutoff membrane (Millipore Co.). Prior to electrophoresis, protein concentrations were determined using the BCA protein assay.⁴⁵

Synchrotron Radiolysis. Synchrotron radiolysis was performed as described previously.⁴⁰ Radiolysis was performed on the XLRM2 beamline at The J. Bennett Johnston, Sr. Center for Advanced Microstructures & Devices (CAMD) synchrotron. PS II membranes (200 μL at 2 mg of chl/mL) were exposed for various lengths of time at room temperature in a multichannel Plexiglas chamber. After exposure, the samples were immediately removed from the chamber and held on ice until being stored at -80°C .

Electrophoresis and Protein Digestion. For the EDC cross-linking experiments, the protein samples were resolved on 12.5 to 20% LiDS–PAGE gradient gels⁴⁶ using ammonium persulfate and tetramethylethylenediamine for polymerization. For the radiolytic experiments, however, the proteins were separated on 12.5 to 20% acrylamide LiDS–PAGE gradient gels using a nonoxidizing system in which the gels were polymerized with riboflavin (in the presence of diphenyliodonium chloride and toluene sulfinate) by exposure to UV light.^{47,48} The upper reservoir also contained thioglycolate. This was required, as standard PAGE polymerization conditions are known to introduce numerous protein oxidation artifacts.⁴⁹ Earlier experiments indicated that PS II proteins resolved in the nonoxidizing gel system exhibited much lower levels of artifactual protein oxidation than when resolved using standard LiDS–PAGE (see Figure S1 of ref 48).

After electrophoresis, the gels were stained with Coomassie Blue and destained, and protein bands of interest were excised. The protein bands were processed for protease digestion (trypsin or trypsin with Lys-C) using standard protocols. After digestion, the proteolytic peptides were processed using a C18 ZipTip prior to mass analysis.

Mass Spectrometry. The proteolytic peptides were resolved using reversed-phase chromatography with mass spectrometry

being performed on a Thermo Scientific LTQ-FT instrument.⁴⁸ This is a hybrid instrument consisting of a linear ion trap coupled to a Fourier transform ion cyclotron resonance mass spectrometer.

Identification and analysis of peptides containing cross-linked products or oxidative mass modifications were performed using the MassMatrix online search engine.^{50,51} A FASTA library containing PsbO, PsbP, and PsbQ proteins was searched, as was a decoy library containing the same proteins but with reversed amino acid sequences. For the identification of cross-linked products, peptides were selected if their *P* value was ≤ 0.0001 . In this study, as expected, no cross-linked products involving the PsbP and PsbQ proteins were identified. Additionally, cross-linked products were identified with StavroX version 3.4.12⁵² using the same FASTA library. This program reports a non-probabilistic score and a false discovery rate (FDR) for each identified cross-linked product. The FDR corresponded to 5%. Only putative cross-linked products that exhibited scores of >100 (which was considered a “significant” score⁵²) and also exhibited scores >3 times the FDR were considered in our study. In both MassMatrix and StavroX, a precursor ion precision of ≤ 5.0 ppm was required.

For the radiolytic footprinting studies, MassMatrix was programmed to search for all of the possible oxidative modifications for the 18 modifiable and identifiable amino acids, excluding glycine and alanine.^{36,53} For the identification of oxidative modifications, a more stringent *P* value (≤ 0.00001) was used. We feel that given the smaller mass modifications introduced by oxidative labeling (2–48 au), the use of a more stringent *P* value was prudent. For both the protein cross-linking studies and the radiolytic modification experiments, the peptides were required to exhibit 0% hits to the decoy library for further consideration.

Protein Modeling. Clustal Omega⁵⁴ was used to align the spinach and *Thermosynechococcus vulcanus* PsbO protein sequences. On the basis of this alignment, the spinach sequence was threaded using SWISS-MODEL⁵⁵ with the structure of *T. vulcanus* PsbO [contained in Protein Data Bank (PDB) entry 3WU2, which is the update of PDB entry 3ARC⁴] as a template. Secondary structure prediction for higher-plant PsbO was performed using Genesilico Metaserver.⁵⁶ The cross-linking distance constraints imposed by the 20 identified EDC cross-linked products were incorporated into the threaded structure using MODELLER.⁵⁷ EDC generates an amide bond between primary amino groups and carboxylates that are in van der Waals contact.^{58,59} To model these interactions, a C_{α} – C_{α} distance of 12.1 Å^{60,61} was used. This distance conservatively takes into account the length of the amino acid R groups and dynamic variations in the protein structure. The 10 lowest-energy models were analyzed by Ramachandran analysis using PROCHECK⁶² and RAMPAGE.⁶³

Antibodies. The antiserum against PsbO was produced using a synthesized peptide consisting of the N-terminal 27 amino acid residues of the mature spinach protein coupled to keyhole limpet hemocyanin. The antiserum was produced and isolated from rabbits after their immunization by standard protocols.⁶⁴ The specificity of the antiserum was confirmed by its specific reaction with the antigenic peptide, isolated PsbO, PS II membrane preparations, and thylakoid membranes.

RESULTS AND DISCUSSION

Protein Cross-Linking. Figure 1 illustrates the results obtained upon treatment of PS II membranes with the cross-linker EDC followed by LiDS–PAGE and immunoblotting.

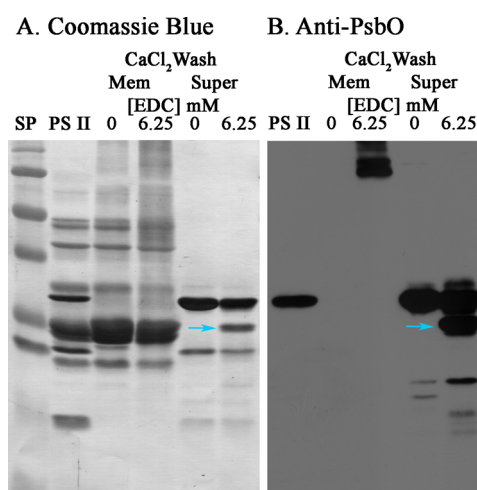


Figure 1. EDC cross-linking of PsbO within the PSII complex. Protein cross-linking was performed as described in [Materials and Methods](#). Illustrated are “Western” blots from the extracts of 1.0 M CaCl_2 -washed PSII membranes (which were obtained after initially washing with 1.0 M NaCl) cross-linked with EDC. Panel A was stained with Coomassie blue, and panel B was probed with anti-PsbO. Intact PS II membranes are labeled PS II and are included for reference. The putative PsbO intramolecular cross-linked product is indicated by the cyan arrows and has an apparent molecular mass of 25 kDa. This protein is the subject of our cross-linking study.

After protein cross-linking, the PS II membranes were washed with NaCl to remove the PsbP and PsbQ proteins. These membranes were then washed with CaCl_2 to remove both unmodified PsbO and PsbO containing putative intramolecular cross-linked residues. This sample was then dialyzed and concentrated by ultrafiltration. As expected, in the absence of EDC, no cross-linked products were observed for PsbO. After cross-linking with EDC, a variety of cross-linked products containing PsbO were observed. Some of these remained associated with the PS II membranes after sequential washing with 1.0 M NaCl, which removes PsbP and PsbQ from the PS II membranes [no cross-linked products containing either PsbP or PsbQ were observed (data not shown)], and then washing with 1.0 M CaCl_2 . These high-apparent molecular mass components represent intermolecular cross-linked products containing PsbO and intrinsic membrane protein components^{65,66} and are the object of continuing investigations. The CaCl_2 treatment removed un-cross-linked PsbO, which migrated at an apparent molecular mass of 29.6 kDa, and PsbO, which contained putative intramolecular cross-links, as indicated by more rapid migration via LiDS–PAGE, exhibiting an apparent molecular mass of 25 kDa. This protein band reacted only with anti-PsbO and not anti-PsbP or PsbQ (data not shown). This 25 kDa protein is the object of this work.

The observed 25 kDa band could be the result of either proteolysis or intrachain protein cross-linking. Subsequent tandem mass spectrometry indicated no proteolysis was evident as we obtained 100% coverage for PsbO, with complete coverage of the N- and C-termini. Consequently, the 25 kDa band appears to result from internal cross-linking with EDC.

In-gel proteolysis followed by tandem mass spectrometry allowed the identification of the cross-linked residues in this 25 kDa band (Table 1). Twenty intrachain cross-linked products were identified within PsbO. All of the cross-linked peptides identified exhibited low *P* values (the probability that the peptide match is a random occurrence) ranging from 1×10^{-4} to

Table 1. EDC-Cross-Linked Residues Identified in PS II-Bound Spinach PsbO^a

higher-plant PsbO cross-linked residues	precursor mass precision (ppm)	StavroX ^b score	FDR ^c	MassMatrix ^d P value	crystal structure distance ^e (C _α -C _α) (Å)
⁴ K- ³² E: ¹ EGG[K] ⁷ R ²¹ GTGTANQCPTV[E]GGVDSFAFKPG ⁴⁴ K	-2.1	-	-	5.0 × 10 ⁻⁵	NA ^f
⁴ K- ³⁶ D: ¹ EGG[K] ⁷ R ²¹ GTGTANQCPTVEGGV[D]SFAFKPG ⁴⁴ K	-3.7	-	-	6.3 × 10 ⁻⁵	NA ^f
⁹ D- ¹⁴ K: ⁶ LT[Y][D]EIQS[K]TYLEV ²⁰ K	+0.7	172	52	5.0 × 10 ⁻⁷	NA ^f
¹⁰ E- ¹⁴ K: ⁶ LT[Y][D]EIQS[K]TYLEV ²⁰ K	+0.3	182	52	3.1 × 10 ⁻⁵	NA ^f
¹⁴ K- ¹⁸ E: ⁶ LT[Y][D]EIQS[K]TYL[E]V ²⁰ K	-0.9	253	39	2.5 × 10 ⁻⁷	8.1
⁶⁰ K- ⁶² E: ⁵⁷ FAV[K]A[E]GIS ⁶⁶ K	0.0	-	-	1.0 × 10 ⁻⁸	5.8
⁶⁰ K- ²³⁴ D: ⁵⁷ FAV[K]AEGIS ⁶⁶ K ²³⁴ [D]V ²³⁶ K	+0.5	278	39	2.5 × 10 ⁻⁹	6.9
⁶² E- ⁶⁶ K: ⁶¹ A[E]GIS[K]NSGPDFQNT ⁶⁶ K	+1.0	-	-	1.2 × 10 ⁻⁸	11.5
⁶² E- ²³³ K: ⁶¹ A[E]GIS ⁶⁶ K ²³¹ VP[K]DV ²³⁶ K	0.0	-	-	5.0 × 10 ⁻⁷	9.9
⁶⁶ K- ⁷¹ D: ⁶¹ AEGIS[K]NSGP[D]FQNT ⁶⁶ K	-2.2	-	-	6.3 × 10 ⁻⁷	10.8
⁶⁶ K- ²³⁴ D: ⁶¹ AEGIS[K]NSGPDFQNT ⁷⁶ K ²³⁴ [D]V ²³⁶ K	-2.6	-	-	5.0 × 10 ⁻⁹	16.1
⁹⁷ D- ¹⁰¹ K: ⁸¹ LT[Y]TLDEIEGPFVSS[D]GTV[K]FEE ¹⁰⁵ K	-4.8	183	59	6.3 × 10 ⁻⁸	9.5
⁹⁷ D- ¹⁰⁵ K: ⁸¹ LT[Y]TLDEIEGPFVSS[D]GTV ¹⁰¹ K ¹⁰² FEE[K]DGIDYAAVTVQLPGGE ¹²² R	+0.1	264	39	-	22.9
¹⁰¹ K- ¹⁰⁴ E: ⁸¹ LT[Y]TLDEIEGPFVSSDGTV[K]FE[E] ¹⁰⁵ K	-4.8	-	-	1.0 × 10 ⁻¹⁰	9.7
¹⁰¹ K- ¹⁰⁹ D: ⁸¹ LT[Y]TLDEIEGPFVSSDGTV[K]FEE ¹⁰⁵ K ¹⁰⁶ DGI[D]YAAVTVQLPGGE ¹²² R	+0.11	249	39	-	8.6
¹⁰³ E- ¹⁰⁵ K: ¹⁰² F[E][K]DGIDYAAVTVQLPGGE ¹²² R	+0.3	-	-	1.9 × 10 ⁻⁹	6.9
¹⁰⁵ K- ¹⁰⁶ D: ¹⁰² FEE[K][D]GIDYAAVTVQLPGGE ¹²² R	+0.4	-	-	3.1 × 10 ⁻⁶	3.8
¹³⁷ K- ¹³⁹ E: ¹³¹ QLVASG[K]P[E]SFGDFLPSY ¹⁵¹ R	+1.4	207	39	-	9.9
¹⁸⁰ D- ¹⁸⁶ K: ¹⁶² GGSTGYDNAVALPAGGRG[D]EEELQ[K]ENN ¹⁹⁰ K	+1.1	224	59	-	NA ^f
¹⁸² E- ¹⁸⁶ K: ¹⁶² GGSTGYDNAVALPAGGRGDE[E]ELQ[K]ENN ¹⁹⁰ K	+0.2	223	59	1.0 × 10 ⁻⁵	9.7

^aStavroX⁵¹ and MassMatrix⁵² were used to identify the various cross-linked products. ^bA significant score: qualifying cross-links have a StavroX score of >100 and >3 times the FDR. ^cFDR, the StavroX false discovery rate, represents a score at which the rate of false discovery is <5% ^dP value, calculated by MassMatrix using max(pp1,pp2), pptag. ^eDistances between the corresponding residues in the cyanobacterial PsbO structure of PS II. These residues are not necessarily fully conserved in higher-plant PsbO. ^fNot applicable, as no corresponding residues are present.

5 × 10⁻¹⁰. The quality of the data used in this work is illustrated in Figure 2. This figure illustrates the mass analysis for the intrachain cross-link involving ¹⁴K and ¹⁸E, which was the median-P value cross-linked product (P = 2.5 × 10⁻⁷) identified in this study. This peptide exhibits nearly complete y- and b-ion series (Figure 2A,B), the exception being that residues that lie between the two cross-linked residues cannot be identified using MassMatrix. A heat map, which visually assists in the identification of cross-linked products in MassMatrix, is also shown (Figure 2C). This cross-linked species was additionally identified with StavroX (Table 1).

Structure of Higher-Plant PsbO. Table 1 summarizes the cross-linked products identified using MassMatrix and StavroX. For the majority of the cross-linked products observed, analogous residue pairs are present in cyanobacterial PsbO. The C_α-C_α distances between these corresponding residue pairs in cyanobacterial PsbO are generally within 12.1 Å (Table 1); this was expected and highlights the overall structural similarity between the higher-plant and cyanobacterial PsbO proteins. Only two anomalously long distances were observed. In *T. vulcanus*, the residues that correspond to the ⁶⁶K-²³⁴D cross-linked residues in spinach are residues ⁵⁸K and ²³¹E. In the crystal structure, the α-carbons of these residues are 16.1 Å apart. This might indicate a modest conformational difference of ~4 Å between the higher-plant and cyanobacterial proteins. The second anomaly is more difficult to explain. In *T. vulcanus*, the residues that correspond to the ⁹⁷D-¹⁰⁵K cross-linked residues in spinach are residues ⁸⁹D and ⁹⁷E. In the crystal structure, the α-carbons of these residues are 22.9 Å apart. This would indicate a rather large conformational (>10 Å) difference between the higher-plant and cyanobacterial proteins. Interestingly, these residues flank a domain on the PsbO protein that is resistant to

radiolytic modification (see below). At this point in time, we cannot explain this anomaly.

The residues located at the N-terminus of the higher-plant protein (¹E-¹⁰E) are not present in cyanobacterial PsbO. Three cross-linked pairs were present in this domain. Additionally, cross-linked residues were located in the vicinity of spinach residues ¹³⁸P and ¹³⁹E. The eight-residue insertion that forms the cyano loop⁶⁷ in the *T. vulcanus* PsbO structure lies between ¹³⁸P and ¹³⁹E in the higher-plant protein. Finally, cross-linked residue pairs that lie near spinach residues ¹⁷⁸R-¹⁸⁰D that are absent in *T. vulcanus* PsbO were identified. This cross-linking information places constraints on the possible structures that can be assumed by higher-plant PsbO when it is bound to PS II.

Given these distance constraints provided by the observed cross-linked products (Table 1), molecular dynamics refinement can provide useful models for higher-plant PsbO, particularly with respect to its 10 N-terminal amino acid residues, which are not conserved in the cyanobacterial protein. MODELLER⁵⁷ was used to provide the structures shown in Figure 3, which illustrates molecular dynamics structural refinements either in the absence (Figure 3A) or in the presence (Figure 3B) of the EDC-cross-linking distance constraints. In both panels, the 10 lowest-energy structures for PsbO are illustrated in different shades of blue and are aligned with the *T. vulcanus* protein (colored orange). Without inclusion of the distance constraints, the N-terminus is highly disordered. Inclusion of the distance constraints, however, yields a family of very similar structures, all of which exhibit similar low DOPE scores of approximately -19500.⁵⁷

It should be noted that secondary structure analysis using the Genesilico Metaserver⁵⁶ of the N-terminal domain of PsbO (¹E-¹⁰E), which is not present in the cyanobacterial protein, indicates that the consensus secondary structure for this region

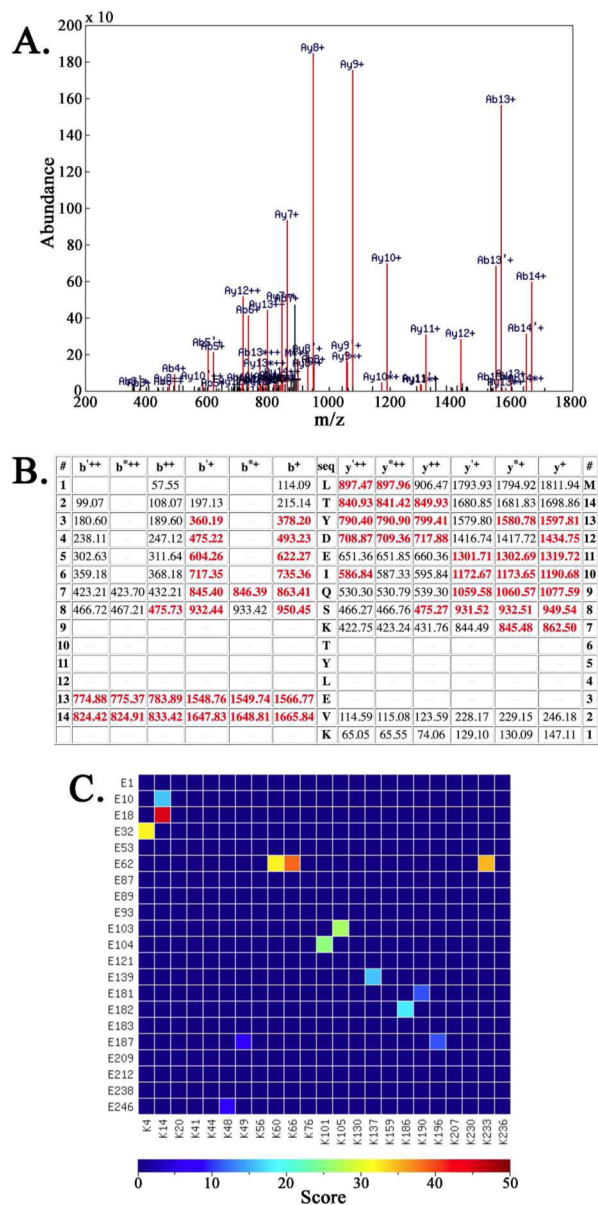


Figure 2. Quality of the mass spectrometry used in this study. Shown are the mass spectrometry data obtained for the median-*P* value cross-linked product ¹⁴K–¹⁸E (Table 1; *P* = 2.5 × 10⁻⁷). (A) Mass spectra obtained for this peptide. Identified ions are colored red and those not identified black. (B) Table of predicted ions from this peptide. Identified ions are colored red and those not identified black. In loop peptides, MassMatrix cannot identify residues lying between the cross-linked residues. Note the nearly complete *y*- and *b*-ion series obtained. Additionally, *y*⁺⁺⁺ and *b*⁺⁺⁺ ions exhibiting neutral loss of water and *y*^{*++} and *b*^{*++} ions exhibiting neutral loss of ammonia are shown. (C) Heat map that assists in the identification of cross-linked species. The identified peptide is colored red.

contains a short α -helix (⁷T–¹⁰E). This program utilizes seven independent secondary structure prediction algorithms. These algorithms uniformly predicted an α -helix at this location, and consequently, this was incorporated into the distance-constrained models.

Elucidation of the structure of the N-terminus of PsbO is critical for understanding the function of this component. Early reports indicated that the 16 N-terminal amino acid residues (¹E–¹⁶Y) were required for binding to PS II and efficient oxygen

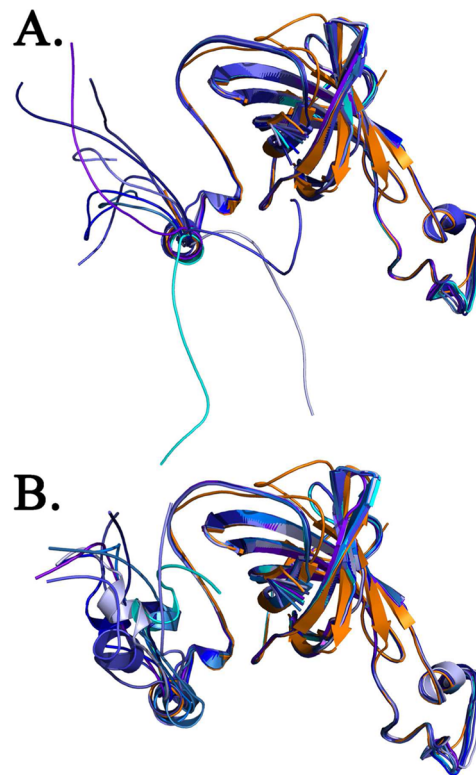


Figure 3. Distance-constrained molecular dynamics refinement for the N-terminus of higher-plant PsbO. Shown are the top 10 models for cases in which (A) no EDC-cross-linking distance constraints are incorporated and (B) EDC distance constraints are incorporated. The individual PsbO models are shown in various shades of blue and are aligned with cyanobacterial PsbO (colored orange). All models exhibited similar low DOPE scores of approximately -19500.

evolution in higher plants.⁶⁸ Subsequently, site-directed mutagenesis studies indicated that two binding determinants for PsbO are located in this N-terminal domain (Figure 4). One determinant (⁴K–¹⁰E) is found in higher plants (and green algae), and its deletion abolishes 50% of the oxygen evolving activity and results in the loss of 50% of the recombinant PsbO binding.⁴¹ The second binding determinant (¹⁵T–¹⁸E) is present in both higher plants and cyanobacteria; its deletion leads to a further loss of PsbO binding and very low oxygen evolution rates.⁴² These authors hypothesized that the deletion of ⁴K–¹⁰E prevented one of two copies of PsbO from efficiently binding to the photosystem. The deletion of ¹⁵T–¹⁸E led to the loss of a second copy. The locations of these two experimentally determined binding determinants are illustrated in Figure 4B. It should be noted that the biochemical evidence that suggests that two copies of the PsbO protein are present per PS II monomer in higher plants is quite strong (for in-depth reviews, see refs 13 and 26).

Radiolytic Footprinting of PsbO. Radiolytic footprinting allows the identification of amino acid residues that are exposed to bulk solvent.^{34–36} The [•]OH that is produced during radiolysis is extremely reactive and can modify 18 different residues that can be identified by mass spectrometry.³⁶ Consequently, radiolytic footprinting is significantly more robust than other chemical modification techniques that can usually label only a few types of residues.^{64,69–71}

Table 2 and Figure 5 present the results from the radiolytic footprinting of the PsbO subunit in association with the PS II complex. Mass spectrometry coverage of the PsbO protein was

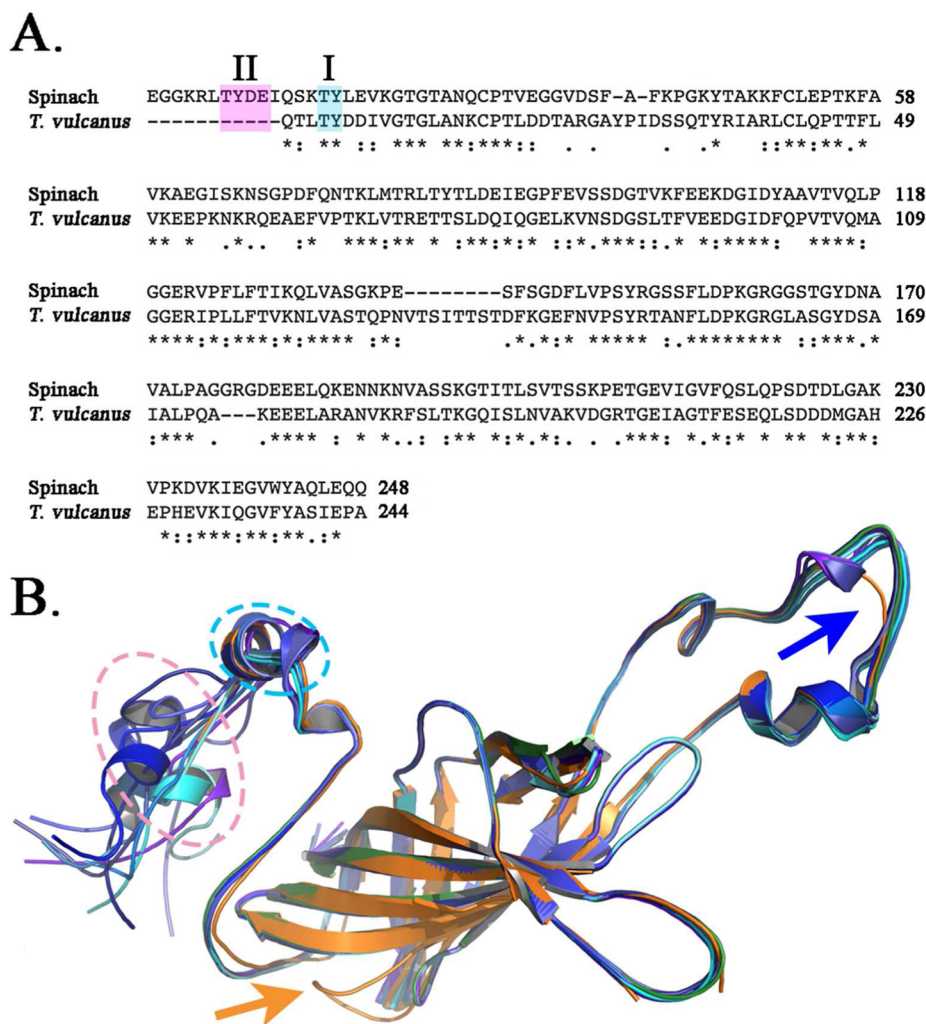


Figure 4. Binding determinants for higher-plant PsbO. (A) Clustal Omega alignment of spinach and *T. vulcanus* PsbO proteins. This alignment is similar to that presented by Popelkova and Yocum.^{41,42} Binding determinant I (highlighted in cyan) is found in all oxygenic organisms, while binding determinant II (highlighted in magenta) is found only in the green plant lineage. (B) Distance-constrained molecular dynamics refinements for spinach PsbO. Shown are the top 10 models of PsbO presented in different shades of blue. These were aligned with cyanobacterial PsbO (orange, PDB entry 3WU2). Binding determinants I and II are outlined in cyan and magenta dashes, respectively. All of the models illustrated fulfill the EDC distance constraints and exhibit similar low DOPE scores (approximately -19500). The orange arrow indicates the location of the cyano loop (*T. vulcanus*, ¹³¹N-¹³⁸S), while the blue arrow indicates the location of the short deletion (spinach, ¹⁷⁸R-¹⁸⁰D) in the cyanobacterial protein.

Table 2. Oxidative Modifications of PsbO^a

⁷T+stcb, ⁸Y+do, ⁹D+gam, ¹⁰E+ca/gam, ¹¹I+go, ¹²Q+ca, ¹³S+stcb, ¹⁴R+go, ¹⁷L+ca/go, ¹⁸E+gam, ¹⁹V+ca, ²⁰K+ca/go, ²²T+stcb, ²⁴T+stcb, ²⁶N+go, ²⁷Q+ca, ²⁹P+ca, ³⁰T+stcb, ³¹V+ca, ³²E+gam, ⁵⁰F+go, ⁵³E+gam, ⁵⁵T+go, ⁵⁶K+ca, ⁵⁷F+go, ⁶⁰K+go, ⁶⁵S+stcb, ⁶⁶K+ca, ⁸⁸I+go, ⁸⁹E+gam, ⁹¹P+ca, ⁹⁶S+go, ⁹⁷D+gam, ⁹⁹T+stcb, ¹⁰⁰V+go, ¹⁰¹K+ca/go, ¹⁰²F+go, ¹⁰³E+ca/gam, ¹⁰⁴E+ca/gam, ¹⁰⁵K+ca/go, ¹⁰⁶D+ca/gam/go, ¹⁰⁸I+ca/go, ¹⁰⁹D+gam/go, ¹¹⁰Y+go, ¹¹⁶Q+ca, ¹¹⁸P+go, ¹²¹E+gam, ¹²²R+ca, ¹³⁸P+ca, ¹³⁹E+gam, ¹⁴⁰S+go/stcb, ¹⁴¹F+do, ¹⁵⁶L+ca, ¹⁵⁷D+gam, ¹⁵⁹R+go, ¹⁶⁵T+stcb, ¹⁶⁷Y+do, ¹⁶⁸D+gam/do, ¹⁷⁴P+ca, ¹⁷⁸R+go, ¹⁸⁰D+gam, ¹⁸¹E+gam, ¹⁸²E+ca, ¹⁸³E+ca, ¹⁸⁴L+gam, ¹⁸⁵Q+go, ¹⁸⁶K+ca, ¹⁸⁷E+gam, ¹⁸⁸N+gam, ²⁰⁷K+ca/go, ²⁰⁸P+ca, ²⁰⁹E+gam, ²¹⁰T+go, ²¹²E+gam, ²²⁶D+gam, ²²⁷L+ca, ²³⁰K+go

^aMassMatrix was used to identify the locations of oxidative modifications. In some instances, different types of oxidative modifications were observed for a given residue. Key: ca, carbonyl addition, +13.98 Da; do, double oxidation, +31.99 Da; go, general oxidation, +15.99 Da; gam, Glu/Asp decarboxylation, +30.01 Da; stcb, Ser/Thr carbonyl, -2.02 Da. It should be noted that while a total of 12 different types of oxidative modifications were incorporated into the MassMatrix searches, only these five types were actually observed.

100% upon integration over all of the irradiation time points (0, 4, 8, and 16 s). Table 2 identifies the residues and types of oxidative modifications observed. The individual time points are not shown; Table 2 and Figure 5 represent a unified set of all of the identified oxidatively modified residues observed in the experiment at all time points.

In Table 2, 77 oxidatively modified PsbO residues were identified. Using modern high-resolution and sensitivity mass spectrometers (FTICR and Orbitrap-class instruments), the oxidative modification of 18 amino acid residues can be detected and identified.^{36,72} Only glycine and alanine remain refractory to radiolytic analysis, because of their low reactivity. As expected, in

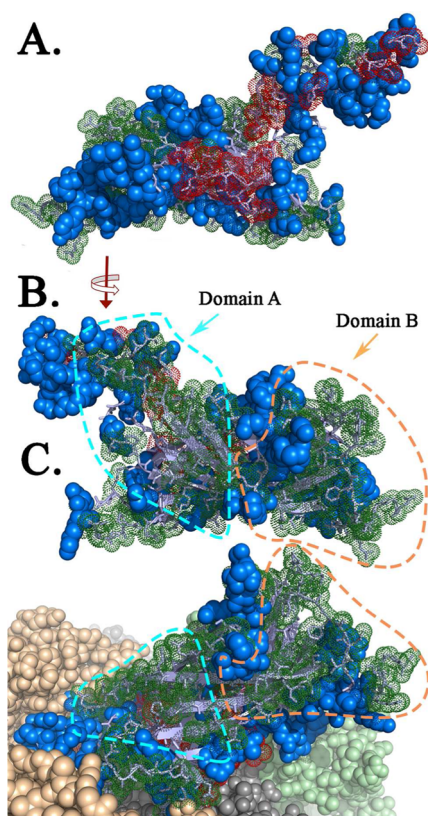


Figure 5. Radiolytic mapping of solvent-exposed domains on higher-plant PsbO. Seventy-seven oxidatively modified residues (blue spheres) were identified. These are listed in Table 2. The mass spectrometry coverage of PsbO was 100%. (A) View of PsbO from the surface of the PS II intrinsic proteins. Residues that were not oxidatively modified and associate with the intrinsic components of the photosystem are shown as red dotted spheres. (B) View of PsbO from the lumen looking down onto PS II. No oxidatively modified residues were identified in the large contiguous lumenally exposed domains labeled domain A (outlined in cyan) and domain B (outlined in orange). Residues that were not oxidatively modified and face the bulk solvent are shown as green dotted spheres. We speculate that these are shielded from oxidative modification by as yet unidentified PS II components. (C) Spinach PsbO shown within the context of cyanobacterial PS II. CP43 is colored green; CP47 is colored brown, and all other subunits are colored gray.

some instances, multiple different oxidative modifications were observed for the same residue.

In Figure 5, the locations of the radiolytic modifications are shown. In Figure 5A, the PsbO protein is viewed from the surface of the intrinsic proteins looking toward the face of PsbO that interacts with the photosystem. In this view, the domains on PsbO that interact with the intrinsic subunits and are shielded from the bulk solvent and, consequently, radiolytic modification are illustrated. Three shielded regions are evident (^{76}K – ^{87}E , ^{149}S – ^{155}F , and ^{169}N – ^{173}L), and these correspond to regions of cyanobacterial PsbO that interact with the intrinsic proteins in the cyanobacterial crystal structure (^{76}K – ^{87}E interacts with CP43 and D1; ^{149}S – ^{155}F interacts with CP43 and D1; ^{169}N – ^{173}L interacts with CP47 and D2) of Umena et al.⁴

In Figure 5B, the protein is viewed from the lumen, looking down onto the exposed surface of the PsbO protein. While numerous exposed residues are modified, a large contiguous domain that is not susceptible to oxidative modification is evident. This includes a large region wrapping around the β -barrel. This is not due to the presence of a high proportion of glycyl and

alanyl amino acid residues, which cannot be identified using this technique. Approximately 80% of the residues in this domain would be identifiable after oxidative modification. This compares favorably to the percentage (85%) that would be identifiable given the standard abundance of amino acids within proteins (Release Notes for UniProtKB/Swiss-Prot data bank, April 2013). In the cyanobacterial crystal structure, these domains do not interact with other PS II subunits, are surface-exposed, and, consequently, should be highly susceptible to radiolytic modification.

The overall pattern of oxidative modifications of higher-plant PsbO is seen within the context of PS II in Figure 5C. Higher-plant PsbO was aligned with the PsbO subunit of *T. vulcanus* as it appears in the cyanobacterial photosystem (PDB entry 3WU2⁴). Because the overall fold of higher-plant PsbO and cyanobacterial PsbO is quite similar (Figure 4B) and because the major intrinsic components of the photosystem that interact with PsbO (D1, D2, CP43, and CP47) are highly conserved, we believe that this model strongly approximates the organization of a single copy of higher-plant PsbO in association with one PS II monomer. What is clear from panels B and C of Figure 5 is that large portions of the higher-plant PsbO protein, which by analogy to the cyanobacterial subunit should be exposed to the bulk solvent, are not observed to be radiolytically modified.

A number of hypotheses could explain this observation. It is formally possible that higher-plant PsbO undergoes a large conformational change upon being associated with PS II, shielding the unmodified residues from the bulk solvent and, consequently, radiolytic modification. In our view, this is highly unlikely and would require unprecedented conformational flexibility of the β -barrel of the protein. Additionally, it would require that the overall architecture of higher-plant PsbO be significantly different from that of the cyanobacterial protein. No evidence of this exists.

A second hypothesis is that the unmodified domains represent regions on the PsbO protein that are shielded by interaction with other PS II subunits. Several possibilities exist. The unmodified PsbO domains could be regions that interact with PsbP and PsbQ. Several investigators have examined the interaction of these subunits with the photosystem. In cyanobacteria, Liu et al.³¹ proposed a binding domain for CyanoQ at the lumenal PS II dimer interface. These authors observed cross-linking between PsbO and CyanoQ that strongly supported their hypothesis. If PsbQ were positioned in an analogous location in higher-plant PS II, this could explain the lack of radiolytic modification in domain A (Figure 5B,C), which lies near the PS II dimer interface. One such model was previously proposed for higher-plant PS II.³³ It should be noted that other investigators have presented alternative models for the location of higher-plant PsbQ, suggesting that it lies at the periphery of PS II and is closely associated with CP43.^{11,12,32,33} In these models, PsbQ was suggested to be located in a position roughly similar to that of PsbQ' in the red alga *C. caldarium*.²⁵ If this positioning is correct for higher-plant PsbQ, then the observed absence of modifications in domain A (Figure 5B,C) cannot be explained. In any event, the presence of unmodified residues in domain B (Figure 5B,C) cannot be attributed to the presence of PsbQ in any of the models that have been presented previously.

It also appears very unlikely that PsbP could occupy domain B. In higher-plant PS II membranes, Ido et al.^{29,32} observed EDC cross-linking between PsbP:¹A and PsbE:⁵⁷E, indicating that these residues are in van der Waals contact. The closest approach between PsbE:⁵⁷E and domain B is >80 Å, which would appear

to preclude the association of PsbP with domain B of PsbO (Figure 5B,C).

Other components have been reported to be associated with PS II membranes and core complexes. These include catalase and polyphenol oxidase,⁷³ TL29,^{74,75} TLP18.3 (Psb32),^{75,76} and a YCF39-like protein.⁷⁵ It is possible that these (and perhaps other) components are associated with PS II membranes but have generally escaped detection, possibly because of the substoichiometric abundance or unusually poor protein staining. The presence of a substoichiometric component in association with domain B (Figure 5B,C), for instance, could reduce the amount of oxidative modification in the domain to below levels that can be detected by mass analysis.

Clearly, one of the most intriguing possibilities is that a second copy of PsbO may be associated with domain B. While the biochemical evidence of the presence of a second copy of PsbO within the PS II monomer is very strong,^{13,26} it must be stressed that no direct structural evidence of a second copy of PsbO has been forthcoming. At this time, we cannot distinguish between these various possibilities.

CONCLUSIONS

In this work, we have provided new information concerning structural features of the PsbO protein from higher plants while it is in association with PS II membranes. The location and organization of the N-terminal 10-amino acid residue extension that is present in higher plants, but not in cyanobacteria, have been examined and structural models for this domain presented. This region appears to be instrumental in the binding of PsbO in higher plants.^{41,42} Additionally, radiolytic footprinting experiments indicate that large contiguous domains on the surface of PsbO are resistant to oxidative modification and, consequently, appear to be shielded from the bulk solvent by as yet unidentified PS II components. Ongoing investigations are underway in an attempt to identify the species responsible for this observation.

NOTE ADDED IN PROOF

Recently, Wei et al. (2016) (Nature DOI: 10.1038/nature18020) have presented a cryo-EM structure of an inactive spinach PS II-LHCII supercomplex at 3.2 Å resolution. The N-terminus of the PsbO protein that is presented in this structure is highly congruent to the structure presented in this communication.

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Notes

The authors declare no competing financial interest.

ABBREVIATIONS

DOPE, discrete optimized protein energy; EDC, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide; LiDS, lithium dodecyl sulfate; PAGE, polyacrylamide gel electrophoresis; PS I, Photosystem I; PS II, Photosystem II.

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